IS Three D power Doppler of the endometrial and sub endometrial regions effective in predicting endometrial implantation? Prospective cohort study

Khalid M Salama MD, Ibrahim I. Souidan MD. Department of Obstetrics and Gynecology, Faculty of Medicine, Benha University, Benha, Egypt.

Abstract

Objective: This study aimed to evaluate the three dimension power Doppler indices together with uterine artery Doppler indices during the day of embryo transfer in predicting the outcome of ICSI cycle.

Study design: One hundred and three healthy women with singleton pregnancy werprospective cohort study.

Patient and methods: One hundred twenty patients were included in the study during ICSI cycles. This work was done at IVF unite of Dar El Teb hospital, Egypt. All patients included in the work had these criteria; age; 22–35 years, BMI; < 35 kg/m², oligo- or oligoasthenospermia. All patients received along agonist protocol of ovarian hyperstimulation and after follicular retrieval; embryos were transferred at the stage of blastocyst. Three D Power Doppler was done at the day of embryo transfer. Quantitative pregnancy tests were done for every patient. The rates of clinical and ongoing pregnancy were estimated. All women were categorized into two categories: with pregnancy and without pregnancy.

Results: Thirty-five percent of patients became pregnant. Our study showed non-significant differences in both groups regarding demographic, clinical and laboratory data except for some vascular parameters (endometrial VI, FI, VFI-subendometrial FI- u PI). Our study revealed a significant increase of some vascular parameters (endometrial VI, FI, VFI-subendometrial FI- u PI) and correlated to pregnancy. The endometrial VFI is the most sensitive vascular parameter correlated to pregnancy.

Conclusion: 3DPD is a useful non invasive predictor for IVF outcome.

Corresponding author:

Khalid M Salama email address: dr.khalidsalama@

gmail.com Postal code: 13617 Address: Qalama- Qaluib

Qalyubia-Egypt.

Telephone number: 01225861026 Key words: 3DPD, Endometrial, subendometrial, implantation NCT04081870 -Khalid 6 dr.khalidsalama@gmail.com

INTRODUCTION

Many factors determine the success in the cycle of IVF/ET; few of them are known to be directly related to the successful outcome. The implantation of good quality embryos remains a rate limiting step in IVF/ET management. In spite of the advances in ovarian stimulation protocols, improvement in culture conditions and the method of assisted fertilization, the implantation rate remains low. The success of embryo implantation depends on a dialogue between the transferred embryo and the receptive endometrium (1).

The receptivity of endometrium was investigated by different strategies like the histological dating of an endometrial biopsy (2), intra-uterine flushing for detection of endometrial cytokines (3), the genomic study of a timed endometrial sample(4). Nevertheless, ERA requires an invasive method, and high cost. Ultrasound can evaluate changes in the endometrium during stimulated cycles by non-invasive technique (5).

The uterine receptivity are controlled by many variables like the endometrial, and the sub-endometrial perfusion (6, 7). Many studies observed a positive correlation between the characteristics of endometrium, and implantation rate after IVF/ICSI cycles, and the poor uterine receptivity was related to impaired blood flow in endometrial and sub-endometrial regions (8, 9, 10).

Ultrasonography was used as a non invasive tool to measure the endometrial thickness to show the effect of endometrial thickness on embryo implantation and endometrial receptivity but unfortunately conflicting findings were obtained. (11).

Some tried to assess the flow of blood in the uterine arteries by Doppler US and they found that uterine arteries Doppler did not represent the actual blood flow in the endometriam. Others tried to use three-dimensional power Doppler ultrasound for measurements of endometrial and sub-endometrial blood flows (1). The endometrial receptivity was evaluated in the endometrial and sub-endometrial blood supplies, especially in intrauterine insemination and IVF-ET cycles (12). This study aimed to investigate the three dimensional power Doppler indices together with uterine artery Doppler indices at the day of embryo transfer in predicting the outcome of ICSI cycle.

Patient and methods:

This prospective cohort research was done at IVF unit of Dar El Teb, Dokki, Egypt, since January 2015 till September 2019. Before the conduction of the study, the Local Ethical Committee approved the work. All women gave consent to participate in the work. One hundred twenty couples included in the work had these criteria; age; 22–35 years, BMI; < 35 kg/m2, male factor with oligoor oligoasthenospermia. Exclusion criteria; -gross pathology in the uterus and tube, -Development

of OHSS, -inadequate response to super ovulation, -failure of mature ovum to fertilize or inadequate development of the embryos to the stage of blastocyst and - If the couple refused to be included in the work at any stage of the treatment cycle.

All patients received long protocol for controlled ovarian overstimulation as described by Chang et al (13). The LHRH agonist ampoules were commenced in the prior mid-luteal phase (decapeptyl R 0.1mg, Triptorelin-Acetate, Ferring GmbH, Wittland 11, D-24109, and Kiel, Germany). After the pituitary down regulation was confirmed, the r FSH vials were given by 225 IU/day (Gonapure 75 IU, IBSA Institute Biochimique SA, Switzerland). During the follow up period of hyperstimulation, dosages were scheduled regarding the response of every woman.

When at least three dominant follicles (a size 18-20 mm) were reached in every patient, the HCG 10000 IU (Epifassi 5000 IU, Epico, Egypt) was taken. The follicles were retrieved 35 hours following HCG administration. Dydrogesterone 30 mg daily (Duphaston, Dydrogesterone 10 mg, Abbott, Pentapharma, Egypt) was used to support the luteal phase. At the day of embryo transfer (blastocyst stage), every woman underwent 3 D power Doppler US. Serum pregnancy test was done after twelve days later to embryo transfer, and if positive (chemical pregnancy), the TVS was used to detect clinical pregnancy while the ongoing pregnancy was detected at the end of first trimester.

Technique of 3D Power Doppler:

We used dedicated 3D transducers to obtain 3D US image. Firstly, determination of the volume box. Secondly, activation of the 3D probe while it was held stationary. Thirdly, the volume data were presented in multi planner display. By using 3D Power-Doppler ultrasound and the VOCAL program (the rotation angel was 30° in our study), we can evaluate the tissue vascularity. Three vascular parameters were used: the Vascularization Index (VI) represents the number of the blood vessels inside the volume box. Flow Index (FI) represents the intensity of blood flow within the area of interest. The Vascular-Flow Index (VFI) represents the number of the blood vessels and the intensity of blood flow within the area of interest. (14) (Figure 1). The "shell" function was used at different

thickness around the predetermined endometrium (in this study, it is estimated to be 5 mm) to measure the sub endometrial volume and estimate the vascularization in this region". (Figure 2)

Statistical analysis

Calculation of the sample size was done by using Open Epi (version 3, open source calculator-SSProor) depending on the number of patients fulfilling inclusion criteria in 6 months and attending to IVF unit of Dar El Teb hospital was estimated by 185 patients and percentage of ICSI success in a prior research is 32.4%(15), so to obtain a research power 80% and CI 95%, at least 120 patients must be included in the research. The variables were presented as mean ±SD. Independent t-test, Mann-Whitney-test and ROC curve analysis were used for statistical analysis. The SPSS program Version 18 was used. The statistical significance was considered when P value <0.05.

Results

Nine patients were excluded from the work from 129 women participating in the study due to; development of OHSS in 2 patients, 2 patients with inadequate ovarian response, failure of mature ovum to fertilize in 3 patients or inadequate development of the embryos to the stage of blastocyst in 2 patients. Figure 1

All included patients had demographic, clinical and laboratory data as presented in table 1. On assessment of serum pregnancy tests, it was found the chemical pregnancy was 39.2% while clinical pregnancy rate was 35%. Only 11 patients had abortions at 7-12 weeks gestation (ongoing pregnancy rate =25.8%). The included women were subdivided into two groups; group A (with pregnancy) and group B (with non pregnancy). Table 2. The no significant differences in both groups regarding the demographic data were presented in Table 1

Assessment of endometrial-sub endometrial and uterine blood flow at the day of embryo transfer, revealed significant increase of endometrial vascularity(VI,FI,VFI) in the pregnant women, significant increase of sub endometrial blood flow (FI) in the pregnant women. Also significant increase of pulsatility index of uterine artery correlated with

pregnancy. The endometrial volume was comparable in both groups. Table 1

At the day of embryo transfer,3 D power Doppler of the endometrial and sub endometrial areas and uterine artery Doppler were used to predict endometrial implantation by The ROC curve analysis. The areas under the curve for E VFI, u PI,E FI, SE FI and E VI were 0.82, 0.75. 0.66, 0.65, and 0.62 at a cut off \geq 0.96 & \leq 1.5, \geq 26.7, \geq 19.9, \geq 19.8, \geq 4.7 indicating that the endometrial VFI is more sensitive and specific than other vascular parameters. Table 3

In other words, the endometrial VFI is the most sensitive endometrial vascular parameter (figure 3), while the sub endometrial FI is the only sensitive sub endometrial vascular parameter (figure 4)detected by 3D PD US. On evaluation of the colour Doppler of uterine artery, it was found that only the u PI was significantly correlated to pregnancy. (Figure 5)

Discussion:

The 3D PD-US angiography is the most important diagnostic tool to evaluate restricted tissue, by showing and calculating relevant parameters (16, 17, and 18). The power Doppler ultrasound has extreme sensitivity to slight blood flow to detect overlapping vessels (19). The restricted tissue like endometrium is important for uterine receptivity (18). Our study showed that the thickness and volume of endometrium were not correlated with pregnancy.

The endometrial thickness was investigated for several years for detection of its relation to pregnancy and the results were controversial because the endometrial thickness was affected by different factors like mechanical stimulation or by the ovarian stimulation treatment method (18). For some times, the endometrial volume was considered an important index for endometrial receptivity with reporting it should be at least 2.0–2.5 ml for establishing pregnancy (20) while another research showed no pregnancy with volume less than 1ml (21). Our study, similar to several studies (22, 23, and 24), found no relation between the endometrial volume together with the endometrial thickness and pregnancy while others concluded a positive

correlation with pregnancy (17). This controversy can be seen in other view as the impact of endometrial thickness or volume on the pregnancy was favored by endometrial vascularization that should be investigated (25, 26). Because sufficient blood supply is necessary for endometrial receptivity (24), the endometrial neomicrovascularisation increases significantly in the follicular and early luteal phase (27) and is affected by different factors like age, medication, hormones (23). The endometrial neomicrovascularisation can be assessed by power Doppler combined with 3D US(17). Vascular indices like VI, FI, and VFI, can be estimated from the total number of color voxels and intensity of blood flow (28). Our study, similar to Wang et al (29) and Singh et al (1), found a positive correlation with pregnancy regarding endometrial vascular indices and sub endometrial flow index, in spite of the assessment of vascularization was questioned in different reports (30, 31,32). The clinical value of 3D-PD US has been intensively studied. Jinno et al (33) stated that the endometrial blood flow during second half of the cycle could predict the outcome in IVF cycles. Ng et al (23) found a positive correlation between flow of blood in endometrial and sub endometrial regions and pregnancy outcome. Furthermore there was a positive correlation between flow of blood in sub endometrial region and some cytokines like IL 15(34) and IL 18 and IL 18 B P (35). Our study, similar to Kim et al (16), found that the endometrial VFI was the most sensitive vascular parameter in predicting pregnancy (0.8) so it is included, recently, in Ultrasound multimodal score to assess the endometrial receptivity. The total score was 18, the lower the score, the worse the endometrial receptivity, and vice versa (36). Based on these results, it is reasonable to hypothesize that the endometrial vascularity is correlated to embryo implantation.

Othman et al (37) stated that the blood flow, in the endometrial and sub endometrial tissue, differed significantly according to BMI. It was lower in obese and overweight women. In obese women, the relatively hyperestogenemia may have negative impact on the receptivity of the endometrium (38). Also the relatively hyper insulinemia decreases the glycodelin level that reduces the fertility at the level of endometriam. (39, 40)

Numerous studies have evaluated the value of measurement of vascularity in endometrial and sub endometrial regions in predicting IVF outcome, at the day of embryo transfer and the day of HCG administration but the results were conflicting (19, 24, 41, 42, 43, 44, 45). These conflicting results and the variations of day timing of measurement explained why still no consensus as to when these measurements should be done (24).

Different thickness of sub endometrial shell was used to assess the sub endometrial vascularity. Some used 2mm (24) and5mm (19) while others used 1mm (22, 23). Our study used 5mm shell. The difference of thickness of subendometrial shell between different studies explained the variation of measurements.

This study was limited as it included only women received long agonist protocol and shell of 5 mm in the sub endometrial region. Our protocol of COS cannot be generalized to other protocols of IVF. Also, this study did not compare other rFSH types.

Conclusion

The use of 3DPD to predict embryo implantation in IVF is of significant value.

Abbreviations

Ovarian hyperstimulation syndrome (OHSS), Estradiol (E2), gonadotropin-releasing hormone agonist (GnRH-a), Luteinizing hormone (LH), in-vitro fertilization (IVF), intra-cytoplasmic sperm injection (ICSI), trans vaginal ultrasound (TVS), human chorionic gonadotropin (hCG), interleukin (IL), embryo transfer (ET), assisted reproductive technology (ART), percent (%),kilogram per square meter Kg/m2 (Kg/m2), recombinant follicle stimulating hormone (rFSH), milli international unit (mIU), milligram (mg), millimeter (mm), number (NO), SD (standard deviation), metaphase 2 (M11) Odd Ratio (OR) confidence interval(CI) , region of interest (ROI), virtual organ computer aided Analysis(VOCAL), binding protein (BP), controlled ovarian stimulation (COS) ,Three dimensions Power Doppler Ultrasound(3 DPD US) uterine artery pulsatility index (u PI), Vascularization Index (VI), Flow Index (FI) Vascular-Flow

Index (VFI),endometrial(E) subendometrial(SE),Endometrial Receptivity Array(ERA).

Declarations

Acknowledgments

Authors would like to thank our nursing staff who provided help for completion of data collection

Funding

The author(s) received no financial support for the research, authorship, and/or publication of this article.

Availability of data and materials

The datasets used and/or analyzed during the current study were available from the corresponding author on reasonable request.

Authors' contributions

KMS: Analysis, Manuscript Drafting, Acquisition of data, Critical Discussion, Management and Follow up of cases; IIS: Study Design, Manuscript Drafting, Acquisition and interpretation of data, Management and Follow up of cases;. Both authors read and approved the final manuscript.

Competing interests

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Consent for publication

Not applicable.

Ethics approval and consent to participate

The study was approved by the Local Ethical Committee of Benha University Hospital and written informed consent was obtained from each participant before the study.

References

- 1. Singh N, Bahadur A, Mittal S, Malhotra N, Bahtt A. Predictive value of endometrial thickness, pattern and sub-endometrial blood flows on the day of hCG by 2D Doppler in vitro fertilization cycles: a prospective clinical study from a tertiary care unit. J Hum Reprod Sci. 2011; 4(1):29–33.
- 2. Noyes RW, Hertig AT and Rock J. Dating the endometrial biopsy. Fertil Steril. 1950; 1:3-25.
- 3. Lédéé-Bataille N, Laprée- Delage G, Taupin JL, Dubanchet S, Frydman R, Chaouat G. Concentration of leukemia inhibitory factor (LIF) in uterine flushing fluid is highly predictive of embryo implantation. Hum Reprod. 2002; 17:213–8.
- Horcajadas JA, Pellicer A, Simón C. Wide genomic analysis of human endometrial receptivity: New times, new opportunities. Hum Reprod Update. 2007; 13:77–86.
- Silva Martins R, Helio Oliani A, Vaz Oliani D and Martinez de Oliveira J. Sub endometrial resistance and pulsatility index assessment of endometrial receptivity in assisted reproductive technology cycles. Reprod Biol Endocrinol..2019; 17:62 https://doi.org/10.1186/ s12958-019-0507-6
- Barker MA, Boehnlein LM, Kovacs P and Lindheim SR. Follicular and luteal phase endometrial thickness and echogenic pattern and pregnancy outcome in oocyte donation cycles. J Assist Reprod Genet. 2009; 26(5):243-249.
- Ng EH, Chan CC, Tang OS, Yeung WS and Ho PC. The role of endometrial and sub-endometrial blood flows measured by three-dimensional power Doppler ultrasound in the prediction of pregnancy during IVF treatment. Hum Reprod. 2006; 21(1):164-70.
- Al-Ghamdi A, Coskun S, Al-Hassan S, Al-Rejjal R and Awartani K. The correlation between endometrial thickness and outcome of in vitro fertilization and embryo transfer (IVF-ET) outcome. Reprod Biol Endocrinol. 2008; 6:37. Doi: 10.1186/1477-7827-6-37.
- Ho M, Huang LC, Chang YY, Chen HY, Chang WC, Yang TC, et al. Electro acupuncture reduces uterine artery blood flow impedance in infertile women. Taiwan J Obstet Gynecol.

- 2009; 48(2):148-51.
- 10. Takasaki A, Tamura H, Miwa I, Taketani T, Shimamura K. et al. Endometrial growth and uterine blood flow: a pilot study for improving endometrial thickness in the patients with a thin endometrium. Fertil Steril. 2010; 93(6):1851-8
- 11. Gingold JA, Lee JA, Jorge Rodriguez-Purata BA, Michael Whitehouse BA, Sandler B, et al. Endometrial Pattern but not Endometrial Thickness Impacts Implantation Rates in Euploid Embryo Transfers .Fertil Steril .2015;104(3):620-628.
- 12. Revel A. Defective endometrial receptivity Fertil Steril: 2012;97:1028-32
- 13. Chang SY, Lee CL, Wang ML, Hu ML, Lai YM, et al. No detrimental effects in delaying initiation of gonadotrophin administration after pituitary desensitization with gonadotrophin-releasing hormone agonist. Fertil. Steril. 1993; 59:183–6.
- 14. Yaman C and Mayer R .Three-dimensional ultrasound as a predictor of pregnancy in patients undergoing ART .J Turkish –German Gynecol Assoc.2012;13:128-34
- 15. Boomsma CM, Kavelaars A, Eijkemans MJC, Lentjes, EG, Fauser BCJM, et al. Endometrial secretion analysis identifies a cytokine profile predictive of pregnancy in IVF. Hum Reprod.2009; 24(6):1427-1435.
- 16. Kim A, Han JE, Yoon TK, Lyu SW, Seok HH, et al. Relationship between endometrial and sub endometrial blood flow measured by three-dimensional power Doppler ultrasound and pregnancy after intrauterine insemination. Fertil Steril.2010; 94:747e52.
- 17. Merce LT, Barco MJ, Bau S and Troyano J. Are endometrial parameters by three dimensional ultrasound and power Doppler angiography related to in vitro fertilization/embryo transfer outcome? Fertil Steril.2008; 89:111e7.
- 18. Zollner U, Specketer M, Dietl J and Zollner K. 3D-endometrial volume and outcome of cryopreserved embryo replacement cycles. Arch Gynecol Obstet 2012; 286:517e23.
- 19. Wu H, Chiang C, Huang H, Chao A, Wang H, et al. Detection of the sub endometrial vascularization flow index by three-dimensional ultrasound may be useful for predicting the

- pregnancy rate for patients undergoing in vitro fertilization-embryo transfer. Fertil Steril.2003; 79:507e11.
- 20. Zollner U, Zollner K, Specketer M, Blissing S, Müller T, et al. Endometrial volume as assessed by three-dimensional ultrasound is a predictor of pregnancy outcome after in vitro fertilization and embryo transfer. Fertil Steril. 2003; 80:1515e7.
- 21. Raga F, Bonilla Musoles F, Casan EM, Klein O and Bonilla F. Assessment of endometrial volume by three-dimensional ultrasound prior to embryo transfer: clues to endometrial receptivity. Hum Reprod.1999; 14:2851e4.
- 22. Schild RL, Knoblock C, Dorn C, Fimmers R, Van der Ven H, et al. Endometrial receptivity in an in vitro fertilization program as assessed by spiral artery blood flow, endometrial thickness, endometrial volume, and uterine artery blood flow. Fertil Steril.2001; 75(2):361–6.
- 23. Ng EH, Chan CC, Tang OS, Yeung WS, and Ho PC. The role of endometrial and sub endometrial vascularity measured by three-dimensional power Doppler ultrasound in the prediction of pregnancy during frozen thawed embryo transfer cycles. Hum Reprod. 2006; 21:1612–7
- 24. Mishra VV, Agarwal R, Sharma U, Aggarwal R, Choudhary S, et al. Endometrial and Sub endometrial Vascularity by Three-Dimensional (3D) Power Doppler and Its Correlation with Pregnancy Outcome in Frozen Embryo Transfer (FET) Cycles. JOGI. 2016;66(S1):S521-S527
- 25. Lesny P, Killick SR, Tetlow RL, Manton DJ, Robinson J, et al. Ultrasound evaluation of the uterine zonal anatomy during in vitro fertilization and embryo transfer. Hum Reprod.1999; 14:1593e8.
- 26. Isaacs JD, Wells CS, Williams DB, Odem RR, Gast MJ, et al. Endometrial thickness is a valid monitoring parameter in cycles of ovulation induction with menotropins alone. Fertil Steril. 1996;65:262e6.
- 27. Santi A, Felser RS, Mueller MD, Wunder DM, McKinnon B, et al. Increased endometrial placenta growth factor (PLGF) gene expression in women with successful implantation. Fertil Steril 2011; 96:663e8.
- 28. Pairleitner H, Steiner H, Hasenoehrl G and

- Staudach A. Three-dimensional power Doppler sonography: imaging and quantifying blood flow and vascularization. Ultrasound Obstet Gynecol. 1999; 14:139e43.
- 29. Wang L, Qiao J, Li R, Zhen X, and Liu Z. Role of endometrial blood flow assessment with color Doppler energy in predicting pregnancy outcome of IVF-ET cycles. Reprod Biol Endocrinol. 2010;8: 122-9
- 30. Nandi A, Martins WP, Jayaprakasan K, Clewes JS, Campbell BK, et al. Assessment of endometrial and sub-endometrial blood flow in women undergoing frozen embryo transfer cycles. Reprod Biomed Online. 2014;28:343–351.
- 31. Silvestre L, Martins WP, and Candido-Dos-Reis FJ. Limitations of three dimensional power Doppler angiography in preoperative evaluation of ovarian tumors. J Ovarian Res.2015; 8:47.
- 32. Nastri CO, Ferriani RA, Raine-Fenning N, and Martins WP. Endometrial scratching performed in the non-transfer cycle and outcome of assisted reproduction: a randomized controlled trial. Ultrasound Obstet Gynecol 2013; 42:375–382.
- 33. Jinno M, Ozaki T, Iwashita M, Nakamura Y, Kudo A, et al. Measurement of endometrial tissue blood flow: a novel way to assess uterine receptivity for implantation. Fertil Steril. 2001;76:1168–1174.
- 34. Ledee N, Chaouat G, Serazin V, Lombroso R, Dubanchet S et al. Endometrial vascularity by three-dimensional power Doppler ultrasound and cytokines: a complementary approach to assess uterine receptivity. J Reprod Immunol, 2008; 77:57–62.
- 35. Wulff C, Weigand M, Kreienberg R, and Fraser HM. Angiogenesis during primate placentation in health and disease. Reprod. 2003; 126:569–577.
- 36. Jiao Y, Xue N, Shui X, Yu C and Hu C. Application of ultrasound multimodal score in the assessment of endometrial receptivity in patients with artificial abortion. Insights into Imaging. 2020 https://doi.org/10.1186/s13244-020-0840-5
- 37. Othman ER, Abdullah KS, Abbas AM, Hussein M, El Snosy E, et al. Evaluation of en-

- dometrial and sub endometrial vascularity in obese women with polycystic ovarian disease. MEFSJ. 2018; 23:324-330.
- 38. Erel CT, and Senturk LM. The impact of body mass index on assisted reproduction. Curr Opin Obstet Gynecol. 2009 Jun; 21(3):228-35. doi: 10.1097/GCO.0b013e32832aee96.
- 39. Carrington B, Sacks G, and Regan L, Recurrent miscarriage: path physiology and outcome. Curr Opin Obstet Gynecol.2005;17(6):591-597 doi:10.1097/01.gco.0000194112.86051.26
- 40. Levens ED, Monica C, and Skarulis MC. Assessing the role of endometrial alteration among obese patients undergoing ART. Fertil Steril. 2008;89(6):1606–1608.
- 41. Kupesic S, Bekavac I, Bjelos D, and Kurjak A.. Assessment of endometrial receptivity by transvaginal color Doppler and three-dimensional power Doppler ultrasonography in patients undergoing in vitro fertilization procedures. J Ultrasound Med. 2001; 20(2):125–34.
- 42. Ng EHY, Chan CCW, Tang OS, Yeung WS and Ho PC. Changes in endometrial and sub endometrial blood flow in IVF. Reprod Bio Med Online. 2009;18(2):269–75.
- 43. Zackova' T, Jarvela IY, Tapanainen JS, and Feyereisl J. Assessment of endometrial and ovarian characteristics using three dimensional power Doppler ultrasound to predict response in frozen embryo transfer cycles. Reprod Biol Endocrinol. 2009;25 (7): 151. doi: 10.1186/1477-7827-7-151.
- 44. Check JH, Dietterich C, Lurie D, Choe JK, and Nazari A. The relationship of color and power Doppler ultrasound parameters of pulsatility and resistance indices and sub-endometrial blood flow with endometrial thickness on day prior to progesterone administration and their relationship to clinical pregnancy rate following frozen embryo transfer. Fertil Steril. 2003; 80:S123.
- 45. Wang J, Xia F, ZhouY, Wei X, Zhuang Y, et al.. Endometrial/Sub endometrial Vasculature and Embryo Transfer Outcome. Ultrasound Med.. 2018;37:149–163.

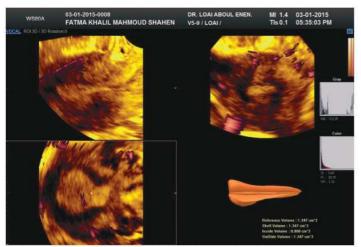


Figure (1): 3 D PD US with VOCAL programme showing endometrial VI, FI, and VFI.



Figure (2):3 D PD US with VOCAL programme showing subendometrial VI, FI, and VFI.

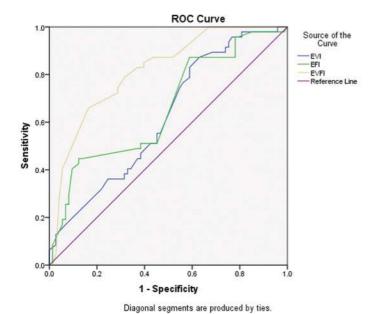
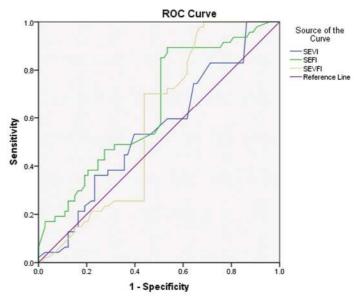


Figure (3):ROC curve of endometrial vascular indices



Diagonal segments are produced by ties.

Figure (4): ROC curve of subendometrial vascular indices

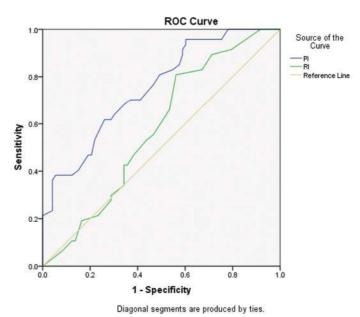


Figure (5):ROC curve of uterine artery Doppler indices

Table 1: shows demographic, clinical and laboratory data and their relations to pregnancy

0 1		3	1	0 3	
Variables	Pregnant (n=47)	Non-pregnant (n=73)	Total (n=120)	t	p
Age (years)	29.87±4.2 (20.0-35.0)	30.38±4.28 (23.0-35.0)	31.52±6.48 (20.0-35.0)	0.64	0.52
BMI (kg/m²)	30.56±3.54 (23.41-38.37)	29.92±2.35 (23.67-39.64)	30.28±3.27 (23.41-39.64)	1.18	0.24
Duration of infertility (year)	5.66 ± 1.56 (3-9)	5.6±2.11 (2-10)	5.79±2.56(2-10)	0.16	0.87
No. of HMG amp	29.94±2.38 (25-37)	30.85±1.84 (26-37)	30.49±2.62 (25-37)	1.89	0.062
Duration of induction : (day)	2.94±1.63 (11-15)	13.32±1.29 (11-15)	13.17±1.44 (11-15)	1.41	0.16
No. of follicles retrieved	15.06±5.08 (9-26)	15.22±4.25 (7-25)	15.16±4.57 (7-26)	0.18	0.86
No. of Metaphase II oocytes:	8.49±2.33 (5-15)	8.7±2.37 (5-14)	8.62±2.35 (5-15)	0.48	0.64
No. of transferred blastocyst:	2.72±0.62 (2-4)	2.68±0.72 (2-4)	2.70±0.68 (2-4)	0.30	0.76
Endometrial VI	4.69±0.89 (0.33-5.52)	4.24±1.28 (0.28-5.51)	4.41±1.16 (0.28-5.52	2.1	0.038*
Endometrial FI	20.21±1.31 (17-22)	19.44±1.22 (17-23)	19.46±1.19 (17-23)	10.72	0.001**
Endometrial VFI	1.1±0.17 (0.86-1.4)	0.90±0.19 (0.06-1.5)	0.98±0.21 (0.06-1.5)	6.26	<0.001**
Endometrial volume	5.62±1.98 (3.572-8.922)	4.94±2.41 (1.26-8.79)	5.35±2.36 (1.26-8.922)	1.6	0.112
Sub VI	2.16±1.71 (0.67-5.93)	1.95±1.78 (0.06-5.86)	2.03±1.75 (0.06-5.93)	0.64	0.53
Sub FI	36.03±20.75 (7.34-65.88)	27.34±20.07 (6.73-65.62)	30.74±20.70 (6.73-65.88)	5.22	0.024*
Sub VFI	1.26±0.67 (0.57-2.48)	1.20±0.98 (0.01-2.53)	0.83±0.78 (0.01-2.53)	0.40	0.69
PI	1.41±0.15 (1.22-1.83)	1.59±0.22 (1.29-1.90)	1.56±0.21 (1.22-1.90)	5.1	<0.001**
RI	0.83±0.06 (0.72-0.92)	0.85±0.07 (0.72-0.95)	0.84±0.07 (0.72-0.95)	1.21	0.23
Endometrial thickness	12.77±0.69 (11.3-13.9)	12.91±0.48 (11.4-14.3)	12.66±0.92 (11.3-14.3)	1.27	0.21
Basal FSH	6.78±0.49 (6.28-7.7)	6.79±0.42 (6.3-7.7)	6.79±0.45 (6.28-7.7)	0.14	0.89
Basal LH	6.07±0.91 (4.9-7.6)	5.92±0.88 (4.8-7.5)	6.09±0.93 (4.8-7.6)	0.87	0.39

Data are presented as mean±SD, and ranges are in parenthesis; *: Significant (p<0.05); **: Highly Significant (p<0.01)

Variables	(n=120)				
variables	No.	%			
Pregnancy test					
-ve	73	60.8			
+ve	47	39.2			
Chemical pregnancy rate	47	39.2			
Clinical pregnancy rate	42	35.0			
Ongoing pregnancy rate	31	25.8			
No. of gestational sac (47)					
1	27	57.4			
2	15	31.9			
3	5	4.2			

Table 3: shows validity of some predictors in prediction of success ICSI.

Variable	Cutoff	AUC	CI	Sens.	Spec.	+PV	-PV	Accu- racy	p-value
EVI	≥4.66	0.62	0.52-0.72	40.4	67.1	44.2	63.6	56.7	0.027*
EFI	≥19.87	0.66	0.56-0.76	51.1	61.6	46.2	66.2	57.5	0.003**
EVFI	≥0.96	0.82	0.74-0.89	74.5	71.2	62.5	81.2	72.5	<0.001**
SEVI	≥1.35	0.55	0.45-0.66	53.2	60.3	46.3	66.7	57.5	0.34
SEFI	≥19.79	0.65	0.55-0.75	55.3	50.7	41.9	63.8	52.5	0.008**
SEVFI	≥1.01	0.58	0.48-0.68	63.8	56.2	48.4	70.7	59.2	0.14
PI	≤1.47	0.751	0.664- 0.838	68.1	65.8	56.1	76.2	66.7	<0.001**
RI	≤0.86	0.578	0.476- 0.68	61.7	49.3	43.9	66.7	54.2	0.15

AUC: Area under curve; CI: Confidence interval; +PV: Positive predictive value; -PV: Negative predictive value; Sens.: Sensitivity; Spec.: Specificity; *: Significant (p<0.05); **: Highly Significant (p<0.01)